INTRODUCTION

Disorders of sex development (DSD) are a group of congenital conditions characterized by atypical development of chromosomal, gonadal or phenotypic sex. Previously these were termed as “Intersex disorders”.1 The most common clinical presentation is variable degree of genital ambiguity in the neonatal period. However, if genital ambiguity is not present or remains unnoticed, they can present later in life with primary amenorrhea, inguinal hernia or virilization in an apparent female, gynaecomastia or delayed puberty in an apparent male, or infertility in both genders.2 Genital ambiguities are reported to occur at an incidence of 1 per 4500 live births.1

DSD arise from abnormalities in gonadal determination or sex differentiation, which are under complex control of various genes and hormones. The chromosomal sex (XX or XY) determines the gonadal sex, which is translated into the phenotypic sex. Under the influence of the sex determining region of the Y chromosome (SRY gene), the undifferentiated, bipotential gonad develops into the testes. Under the action of hormones secreted from testes, the internal ducts and external genitalia differentiate into a male sex phenotype. In the absence of the SRY gene, female sexual differentiation occurs. The bipotential gonad develops into the ovary, and the internal and external genitalia differentiate into that of a female in absence of testicular hormones.3 Downstream of the SRY gene, other genes and transcription factors have important roles in testes development.4 The ovarian development also depends on expression of critical genes. Any error during sex determination or differentiation, results in ambiguous genitalia or DSD.5

The birth of a child with ambiguous genitalia is a medical and social emergency. It is not only a cause of emotional stress for the family but also a diagnostic challenge for the physician.6 Urgent evaluation is necessary to establish a correct diagnosis as soon as possible. The aim should be, to counsel the parents about the treatment options and gender assignment in time to prevent medical and psychological complications. DSD management requires a multi-disciplinary team that should include a pediatric endocrinologist, pediatric urologist, geneticist, and a psychologist.7

Patients should be evaluated by detailed family and antenatal history, thorough physical examination,
karyotyping, radiological imaging, and hormonal testing. Molecular testing for various gene mutations, where available, should also be performed. Patients should be investigated in a tertiary care setup where all these diagnostic facilities are available. The 2006 consensus statement of the ESPE proposed a revised nomenclature and classification of DSD taking into account genetic makeup and descriptive terminologies. The main categories include 46XX DSD, 46XY DSD, and DSD with sex chromosome abnormalities.\(^1\) Cytogenetic analysis to establish the karyotype is thus essential for classification and diagnosis of DSD cases. As the differential diagnosis and further investigations depend on the karyotype, it is the first-line testing in DSDs. It is also essential for excluding mosaicism or other chromosomal abnormalities.\(^8\)

Armed Forces Institute of Pathology (AFIP) is a tertiary care referral centre providing advanced diagnostic facilities like cytogenetic analysis and hormonal testing. Currently there is limited data available in Pakistan about DSD cases. This study was conducted with an aim to determine the frequency of DSD cases presenting to our setup and to classify them according to their chromosomal makeup.

**METHODOLOGY**

This cross-sectional study was conducted at Haematology Department, Armed Forces Institute of Pathology (AFIP), Rawalpindi, from August 2018 to March 2019.

**Inclusion Criteria:** All the patients referred for cytogenetic analysis with the indication of ambiguous genitalia were included in the study.

**Exclusion Criteria:** Patients referred for cytogenetic analysis due to other indications, e.g. primary amenorrhea in females or pubertal delay in males, were excluded.

Sample size was calculated using WHO sample size calculator. Study was conducted after the ethical approval of the institutional review board (IRB no. 18/859). All the patients were included in this study after the informed consent of patients or their parents. Non-probability consecutive sampling technique was used.

All the cases were subjected to a detailed history which included age of presentation, sex of rearing, consanguinity, family history, history of sibling death, maternal exposure to androgens during pregnancy or virilization symptoms in the mother. Criteria for ambiguous genitalia included overt genital ambiguity, apparent female genitalia with enlarged clitoris, posterior labial fusion, or inguinal/labial mass, apparent male genitalia with bilaterally descended testes, hypospadias, or micropenis. Physical examination included, genital examination with stress on presence or absence of testes and their site, and the presence or absence of secondary sexual characteristics in older patients. Patients were also examined for dysmorphic features or other congenital anomalies. Record of radiological investigations was obtained from the patients. These included abdomino-pelvic ultrasonography and/or MRI.

Cytogenetic analysis included chromosomal karyotyping using the conventional G-banding technique. 3ml of heparinized peripheral blood samples were collected. Samples were subjected to 72 hours culture in RPMI 1640 medium at 37-C. Harvesting was done to obtain metaphases by first adding 1% colchicine followed by incubation, centrifugation and addition of hypotonic solution of 1% KCl. Fixation was done by methanol and glacial acetic acid in a ratio of 3:1 after which slides were prepared. Slides were examined under the microscope after Leishman staining to look for presence of at least 20 metaphases that rendered the culture successful. After Giemsa trypsin banding slides were analyzed by CytoVision semi-automated image analysis and capture system. A minimum of 20 metaphases were analyzed and interpreted according to the international system of cytogenetic nomenclature (ISCN).

The results of karyotyping were used to classify patients as having 46XX DSD, 46XY DSD or Sex chromosome DSD. All patients were also tested for 17-hydroxyprogesterone (17-OHP) levels to exclude Congenital Adrenal Hyperplasia (CAH), as advised by their referring physicians. All the information was entered on a predesigned pro forma. The data was analyzed by SPSS version 24. Quantitative variables were presented in terms of median while qualitative variables as frequency and percentages.

**RESULTS**

Fifty one cases of ambiguous genitalia were studied. Their age ranged from 3 days - 30 years with a median age of 15 months. Among them, 29 (56.8%) were in the age group of 0-1 years, 13 (25.4%) were in the age group of 2-6 years while 2 (3.9%) were aged 7-11 years. Four (7.8%) belonged to the age group of 12-16 years and 3 (5.8%) were between 17-30 years as shown in Figure-1. Thirty-three (64.7%) patients had a 46XY karyotype, 17 (33.3%) had a 46XX karyotype while 1 (1.9%) had a 45X/46,XY mosaic karyotype.
Thus they were classified as having 46XY DSD, 46XX DSD and sex chromosome DSD respectively shown in Figure-2.

**Figure-1: Age distribution of study population.**

**Figure-2: Classification of cases of ambiguous genitalia in this series.**

Out of 51 patients, 30 (58.8%) were products of consanguineous marriage. The consanguinity rate was 20 (60.6%) among 46XY DSD and 10 (58.8%) among 46XX DSD cases (Table). Family history of ambiguous genitalia and/or congenital anomalies was found in 8 cases (15.6%).

<table>
<thead>
<tr>
<th>Classification</th>
<th>No. of Cases</th>
<th>Consanguinity</th>
</tr>
</thead>
<tbody>
<tr>
<td>46XY DSD</td>
<td>33</td>
<td>20 (60.6%)</td>
</tr>
<tr>
<td>46XX DSD</td>
<td>17</td>
<td>10 (58.8%)</td>
</tr>
<tr>
<td>Sex chromosome DSD</td>
<td>1</td>
<td>-</td>
</tr>
<tr>
<td>Total</td>
<td>51</td>
<td>30 (58.8%)</td>
</tr>
</tbody>
</table>

Among 46XY DSD cases (n=33), 27 (81.8%) were being reared as males while 6 (18.2%) were being reared as females. Among 46XX DSD cases (n=17), 9 (53%) were being raised as females while 8 (47%) were being raised as males. In the 6 cases with 46XY karyotype being raised as females and the 8 cases of 46XX karyotype being raised as males, karyotyping was repeated to confirm the diagnosis. One case of 45X/46,XY mosaic karyotype was being raised as a female. Its karyotype was also confirmed by repeat testing and examination of 40 metaphases.

Congenital Adrenal Hyperplasia (CAH) was diagnosed in 12 cases (70.5%) of 46XX DSD and in 3 cases (9%) of 46XY DSD based on raised 17-OHP levels. On imaging studies, all patients with 46XX karyotype had internal female genitalia except for 3 (17.6%), who had bilateral undescended testes and no internal female genitalia. No patients with 46XY karyotype had any internal female genitalia. 9 (27.7%) had testes located within scrotal sacs while 13 (39.4%) had bilateral or unilateral undescended testes; both of these groups had normal male internal genitalia. 6 (18.2%) cases had rudimentary testes while 5 (15.1%) cases had absent testes; both of these groups did not have any male internal genitalia. One case of 45X/46,XY karyotype (Turner Mosaic) had female-like ambiguous genitalia with bilateral inguinal testes, however ultrasound revealed an infantile uterus with absent ovaries.

**DISCUSSION**

There is scarce data available on the prevalence and etiologies of ambiguous genitalia/DSD in Pakistan. The process of human sexual development is tightly regulated by various genes and hormones. Any disruption during this process results in disorders of sexual development that have variable presentations and multiple etiologies. They can present at birth as ambiguous external genitalia or may manifest later in life. A newborn with malformed genitalia is a diagnostic and therapeutic challenge for the treating paediatrician. A multidisciplinary team should work closely with the family to establish the correct diagnosis of the abnormality and assign the gender based on the karyotype, endocrine function, fertility potential and surgical options.

In the present study, majority of the patients of ambiguous genitalia presented in infancy 29 (56.8%). This is consistent with published data. A study conducted by Amolo et al. showed that 58% of the cases were less than 1 year of age.

Cytogenetic analysis revealed that the most common karyotype was 46XY 33 (64.7%). Mazen et al. and Al-Agha et al. have also observed a higher incidence of 46XY DSD in their studies, 65.9% and 56.8%, respectively. However, a study conducted by Al-Jurayyan showed that 46XX DSD was more common than 46XY DSD (65.4% and 34.6%, respectively).
Review of published data shows that majority of DSD cases have a normal karyotype i.e. 46XY or 46XX, while karyotype abnormalities leading to sex chromosome DSD are less common.14,15 Our study also showed similar results with only 1 case (1.9%) with a 45, X/46,XY mosaic karyotype.

CAH was diagnosed in 12 cases (70.5%) of 46XX DSD and 3 cases (9%) of 46XY DSD based on raised 17-OHP levels. CAH is the most common cause of DSD worldwide, with an estimated incidence of 1:1400-1:1500 live births; however, it varies among different ethnic groups.16

In the present study, 58.8% of the cases were products of consanguineous marriages. A study from Sudan17 observed a consanguinity rate of 70%. Mazen et al.18 have observed a consanguinity rate of 62.8% among Egyptian patients. Bashamboo and McElreavey18 have suggested that high consanguinity rates may have a role in the higher incidence of DSD cases among certain populations. The incidence of ambiguous genitalia in Arab countries is higher than the incidence reported in studies from European countries.19,20 Arab, African and Asian countries have higher incidence of consanguineous marriages. Consanguinity leads to the accumulation of abnormal recessive genes, and thus results in an increased incidence of conditions manifested by autosomal recessive inheritance. CAH has an autosomal recessive pattern of inheritance, thus leading to its increased incidence in the endogamous communities.21 Few rare conditions of 46XY DSD also have an autosomal recessive inheritance.

Our study showed that a relatively high number of cases of ambiguous genitalia reported to our setup in a short period of time (51 cases in 6 months) as compared to the studies done by Al-Mutair et al.22 (120 patients in 10 years), and Praburam 23 (165 patients in 18 years). This reflects the high burden of DSD in our population. In addition, the frequency is higher than reported by Manzoor et al.24 (300 patients in 7 years) and Raza et al.25 (429 patients in 5 years) in other parts of Pakistan. This may be due to the reason that our cytogenetic lab caters to heavy referrals from all over the country.

Karyotype analysis has a central role in the investigation of DSD. It is essential to classify DSD and helps in narrowing the differential diagnosis and planning subsequent hormonal investigations. Results of karyotyping, radiological imaging and biochemical assays suggest the etiological diagnosis. Early diagnosis is crucial to prevent life threatening complications in case of CAH, to determine the most appropriate sex of rearing, to plan hormonal and/or surgical management, and to counsel the parents and patients.

CONCLUSION

The results of our study indicate that these conditions are not uncommon in our population. Further studies to determine the actual prevalence and various etiologies of DSD cases in Pakistan are required. This will help in creating diagnostic algorithms and management guidelines.

Conflict of Interest: None.

Authors’ Contribution

SAZ: Conception and design of study, collection and analysis of data and drafting of manuscript, AM: Review of manuscript, final approval, RM: Data analysis, Interpretation of data, literature review, AL: Data collection, HMR: Literature review, SF: Manuscript preparation.

REFERENCES