

ACTIVE SURVEILLANCE FOR ASYMPTOMATIC COLONIZATION WITH MULTI-DRUG RESISTANT GRAM-NEGATIVE BACILLI IN PRE-OPERATIVE PAEDIATRIC CONGENITAL CARDIAC DISEASE PATIENTS; PRELIMINARY RESULTS

Sabeen Khurshid Zaidi, Hafsa Khalil, Iftikhar Ahmed, Hajira Akbar, Kamal Saleem, Waqar Ali

Armed Forces Institute of Cardiology/National Institute of Heart Diseases Rawalpindi

ABSTRACT

Objective: To carry out active surveillance for Gram negative Multi drug resistant Bacilli in pre-operative congenital heart disease patients.

Study Design: Prospective cohort study.

Place and Duration of Study: This study was conducted at Department of Paediatric Cardiac Surgery AFIC-NIHD, Rawalpindi from 1st Jan 2015 to 28th Feb 2015.

Methodology: It is an ongoing study, but a preliminary analysis has been conducted on a sample size of 50 patients. Informed consent was taken from patient's parents before sample collection. Samples included throat and rectal swabs which were processed for culture and antibiotic sensitivity of gram negative bacilli isolated.

Results: A total of 14 (28%) patients were identified who were colonized with Multi drug resistant gram negative bacteria (MDR-GNB). Nine isolates were ESBL producers. None of the strains showed Carbapenem resistance. Isolated MDR-GNB were *Escherichia coli* (n=10), *Proteus* species (n=01), *Acinetobacter* species (n=1) and *Klebsiella pneumoniae* (n=02).

Conclusion: The presence of MDR GNB in preoperative paediatric patients addresses the need for antibiotic stewardship programmes and judicious use of antimicrobials in community based settings.

Keywords: Gram negative bacilli (GNB), Multi drug resistance (MDR), Pre-operative.

INTRODUCTION

Multi drug resistant (MDR) gram- negative infections (resistance to three or more than three groups of antibiotics) are associated with increased length of hospital stay, mortality and colossal hospital costs in comparison with infections due to susceptible gram negative bacilli (GNB)^{1,2}. Injudicious antibiotic use has been associated with MDR gram-negative infections³. MDR GNB infection is also associated with resistance of endogenous flora attained as a result of antibiotic pressure or transmission of resistant GNB from the hospital environment⁴. The aim of this study is to determine the frequency of paediatric congenital heart disease pre-operative patients colonized with MDR GNB in our set-up.

PATIENTS AND METHODS

This Prospective cohort study is being conducted at Department of Paediatric Cardiac

Surgery AFIC-NIHD, Rawalpindi in collaboration with Pathology Department AFIC NIHD. Patients undergoing congenital Cardiac Surgery less than 18 years of age are included in this ongoing study. Patients who have length of stay greater than 3 days in current admission were excluded. A preliminary analysis has been conducted on a sample size of 50. The study was approved by the Institutional Ethical and Review Board AFIC/NIHD. Formal approval and informed consent was taken from the parents before sample collection. An especially designed data collection form was used for data collection which included patient demographics, history of prior hospital admission and use of antibiotics. Saline moistened throat and rectal swabs were taken and cultured on blood and MacConkey's agar (Oxoid UK). Gram negative bacilli identified on Gram staining, if yielded were identified using API 20NE (Biomérieux, France). Antimicrobial susceptibility testing was carried out on Mueller-Hinton Agar (Oxoid, UK) by using modified kirby-Bauer disc diffusion technique as per CLSI 2014 criteria⁵. The production of Extended spectrum beta lactamase (ESBL) was

Correspondence: Dr Sabeen Khurshid Zaidi, Department of Pathology, AFIC/NIHD, Rawalpindi
Email: sabeenkhurshid@gmail.com

detected phenotypically by using double disc synergy test using a susceptibility disk (Oxoid UK) containing amoxicillin-clavulanate (20/10

Acinetobacter species (n=01) and Klebsiella pneumoniae (n=02).

Table-1: Correlation of Patients history with MDR-GNB colonization.

History of Patients having colonization of MDR-GNB	MDR-GNB	p-value*
H/o Previous Hosp admission n=10	4(40%)	0.643
Use of Antibiotics in last month n=15	6(40%)	0.243
Use of Antibiotics at the time of admission n=45	18(40%)	0.340
Chest Infection at the time of admission n=7	1(14.2%)	0.357
Fever at the time of admission n=6	2(33.3%)	0.510

*Fisher's exact test

ug), placed in the center of the plate, and disks containing oxyimino-β-lactam the standard ceftazidime (30 ug), ceftriaxone (30 mg), aztreonam (30 mg) or cefpodoxime (10 mg) are placed 20 mm (center to center) from the amoxicillin-clavulanate disk. Plates were then incubated overnight at 35 ± 2°C. Positive result was indicated when the inhibition zones around any of the cephalosporin discs was augmented in the direction of the disc containing clavulanic acid⁶.

The data was entered and analyzed in SPSS version 21 (IBM). Frequencies and percentages were calculated for qualitative variables while mean and standard deviation (SD) were calculated for quantitative variables. Statistical significance between various variables was analyzed using Fisher's Exact Test with a p value of <0.05 considered as statistically significant.

RESULTS

Active surveillance of MDR-GNB is an ongoing study. This is a preliminary analysis of 50 patients who were assessed as per inclusion criteria from 1st Jan 2015 to 28th Feb 2015. The mean age of the patients was 4.5 ± 2.21 yrs. Out of 50 patients 34 (68%) were Male and 16 (32%) were female.

A total of 14 (28%) patients were identified, who were colonized with Multi drug resistant gram negative bacteria (MDR-GNB). As shown in fig no.1, 13 (26%) isolates were rectal and 1(2%) from throat, 9 (64.2%) were ESBL producing all were isolated from rectal swabs. None of the strains showed Carbapenem resistance. Isolated MDR-GNB were Escherichia coli (n=10), Proteus species (n=01),

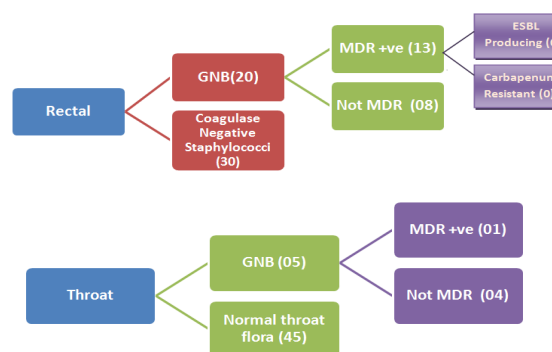


Figure-1: Presence of MDR-GNB in rectal and throat.

Table-2: Antibigram of multi drug resistant gram negative bacteria n=14.

Antibiotics	Resistant	Sensitive
Trimethorprim sulfamethoxazole	11	03
Minocycline	04	10
Gentamycin	06	08
Amikacin	03	11
Ciprofloxacin	07	07
Cefotaxime	05	09
Ceftriaxone	05	09
Colistin	01	13
Imipenum cilastatin	0	14
Meropenum	0	14
Cefoperazone/ Sulbactam	2	12
Piperacillin/ Tazobactam	3	11

10 (20%) Patients had history of previous hospital admissions, out of which 3(30%) were colonized with MDR-GNB. 14% (n=7) and 12% (n=6) had chest infection and fever at the time of admission respectively all of whom were receiving antibiotics at time of admission. 30%

(n=15) patients had history of antibiotic usage prior to congenital cardiac surgery, and 90% (n=45) patients were given antibiotics at the time of admission. Previous history of fever, antibiotic usage, hospital admissions and other infections was not significantly associated with MDR-GNB colonization as shown in Table 1 (P values >0.05). Majority of the MDR GNB were susceptible to Carbapenems and Colistin as shown in Table-2.

DISCUSSION

Patients in pediatric intensive care Units (PICU) are prone to develop infections with MDR organisms. A retrospective cohort study by Siddiqui et al shows increased rates of carbapenem resistant gram negative isolates in children admitted in the intensive care unit⁷. Although all the MDR isolates were resistant to Cephalosporins in our preliminary analysis; so far, none of the isolates are resistant to Carbapenems. Due to increased colonization and infection by multidrug-resistant gram-negative bacilli (MDR GNB), many military hospitals in USA have instituted infection control processes inclusive of active surveillance cultures detection of MDR GNB colonization¹.

Predominantly MDR *Klebsiella pneumoniae*, *Escherichia coli* and *Acinetobacter* species have been implicated in colonization³.

Extended Spectrum β -lactamases (ESBLs) convey resistance to all penicillins, Cephalosporins and Monobactams⁶. More than 50% of the MDR GNB isolates in our study were ESBL producers in contrast to the findings of Villar et al, who detected 18.9% ESBL producers from faecal samples in Argentina which was a community based setting, and 4.9% Carbapenem - resistant GNB were isolated⁸. In another large multicenter study of hospitalized paediatric patients in France, most of the MDR GNB isolates were *Enterobacter cloacae* and *Escherichia coli* with a prevalence of ESBL producing GNB being 5.2% mostly isolated from younger children consistent with our findings⁹. A high prevalence of ESBL GNB colonization has also been reported in Spain¹⁰.

Although this is a preliminary analysis and the sample is limited, no statistically significant association was found with history of previous hospital admission unlike the findings of Patel et al who demonstrated multiple MDR GNB colonizations associated with previous hospital stays in paediatric patients¹¹. The sensitivity of surveillance cultures of ICU patients for MDR GNB in detecting positive clinical cultures for MDR GNB has been reported as 58.8% (95% confidence interval, 48.6 to 68.5%)¹².

The injudicious use of broad-spectrum antibiotics even in community settings paucity of antibiotic stewardship and lack of adherence to antibiotic policies in hospitals has led to increased prevalence of MDR GNB¹³. Major risk factors associated with MDR GNB colonization include older age, hospital admission and antibiotic usage within the last three months and presence of medical devices like Central venous catheters, urinary catheters and mechanical ventilation^{13,14}. The presence of colonization with MDR GNB in this paediatric sample advocates the need for further population based studies to determine the actual burden of MDR GNB in the community. Usage of amoxicillin-clavulanic acid and prolonged hospitalization were significantly associated with carriage of Carbapenemase resistant enterobacteriaceae and majority of the isolates were producing ESBL in Pakistan¹⁵. NDM1 enzyme conferring Carbapenem resistance and a forerunner for pan drug resistance has also been reported from Pakistan; mostly from commensal *Escherichia coli*¹⁶.

CONCLUSION

The presence of MDR GNB in pre-operative paediatric patients addresses the need for antibiotic stewardship programmes, judicious use of antimicrobials in community based settings, hand hygiene compliance and use of risk based contact precautions to prevent cross infection.

Conflict of Interest

This study has no conflict of interest to declare by any author.

REFERENCES

1. S horr AF. Review of studies of the impact on Gram-negative bacterial resistance on outcomes in the intensive care unit. *Crit Care Med*. 2009;37(4):1463-1469.
2. Giske CG, Monnet DL, Cars O, Carmeli Y; ReAct-Action on Antibiotic Resistance. Clinical and economic impact of common multidrug-resistant gram-negative bacilli. *Antimicrob Agents Chemother*. 2008;52(3):813-821.
3. Amy C. Weintrob, MD, Clinton K. Murray et al., Active Surveillance for Asymptomatic Colonization with Multidrug-Resistant Gram-Negative Bacilli among Injured Service Members – A Three-Year Evaluation. *MSMR*. 2013 ; 20(8): 17–22.
4. Weintrob AC, Roediger MP, Barber M, et al. Natural history of colonization with gram-negative multidrug-resistant organisms among hospitalized patients. *Infect Control HospEpidemiol*. 2010; 31(4):330–337. [PubMed: 20175687].
5. CLSI M100-S24 - Performance Standards for Antimicrobial Susceptibility Testing, 2014
6. Rahman M, Rahman MM. Clinical Laboratory and Molecular Detection of Extended Spectrum beta lactamases: A Review Update. *Bangladesh J Infect Dis* 2014;1(1):12-17.
7. Siddiqui NU, Qamar F, Jurair H, Haque A. Multi-drug resistant gram negative infections and use of intravenous polymyxin B in critically ill children of developing country: retrospective cohort study. *BMC Infect Dis*. 2014 Nov 28;14(1):626.
8. Villar HE, Baserni MN, JugoMB. Faecal carriage of ESBL-producing Enterobacteriaceae and carbapenem-resistant Gram-negative bacilli in community settings. 2013 Aug 15;7(8):630-4. doi: 10.3855/jidc.2900.
9. Boutet-Dubois A1, Pantel A, Prère MF, Bellon O, Brieu-Roche N, Lecaillon E et al. Faecal carriage of oxyiminocephalosporin-resistant Enterobacteriaceae among paediatric units in different hospitals in the south of France. *Eur J ClinMicrobiol Infect Dis*. 2013 Aug;32(8):1063-8. doi: 10.1007/s10096-013-1851-7.
10. González-Vélez AE, Díaz-Agero Pérez C, Robustillo-Rodela A, Pita-López MJ, Cornejo-Gutiérrez AM, Pedrero-Pérez P, et al. [Trends in the prevalence of extended-spectrum beta-lactamase-producing gram-negative bacilli in a teaching hospital in Madrid, Spain]. *Med Clin (Barc)*. 2013 Jul 7;141(1):8-12. doi: 10.1016/j.medcli.2012.03.034
11. Patel SJ, O'Toole D, Larson E. A new metric of antibiotic class resistance in gram-negative bacilli isolated from hospitalized children. *Infect Control HospEpidemiol*. 2012 Jun;33(6):602-7. doi: 10.1086/665709. Epub 2012 Apr 13
12. Ridgway JP, Peterson LR, Thomson RB Jr, Miller BA, Wright MO, Schora DM et al. Sensitivity of surveillance testing for multidrug-resistant Gram-negative bacteria in the intensive care unit. *J ClinMicrobiol*. 2014 Nov;52(11):4047-8. doi: 10.1128/JCM.02369-14. Epub 2014 Aug 20.
13. Nordmann P, Cuzon G, Naas T. The real threat of *Klebsiella pneumoniae* carbapenemase-producing bacteria. *Lancet Infect Dis*. 2009;9(4):228-236.
14. Caterino JM. Evaluation and management of geriatric infections in the emergency department. *Emerg Med Clin North Am*. 2008;26(2):319-343, viii.
15. Day KM, Ali S, Mirza IA, Sidjabat HE, Silvey A, Lanyon CV, Cummings SP, Abbasi SA, Raza MW, Paterson DL, Perry JD. Prevalence and molecular characterization of Enterobacteriaceae producing NDM-1 carbapenemase at a military hospital in Pakistan and evaluation of two chromogenic media. *DiagnMicrobiol Infect Dis*. 2013 Feb;75(2):187-91. doi: 10.1016/j.diagmicrobio.2012.11.006. Epub 2012 Dec 14.
16. Sartor AL, Raza MW, Abbasi SA, Day KM, Perry JD, Paterson DL, Sidjabat HE. Molecular epidemiology of NDM-1-producing Enterobacteriaceae and *Acinetobacter baumannii* isolates from Pakistan. *Antimicrob Agents Chemother*. 2014 Sep;58(9):5589-93. doi: 10.1128/AAC.02425-14. Epub 2014 Jun 30.