Crosslinking Salivary Diagnosis with Non-Invasive Insights to Oral Pathology: Novel Systematic Insights to Personalized Medicine in Disease Management

Rehana Kausar, Attia Batool^{*}, Ashfaq Ahmad Shah Bukhari^{**}, Abdul Rehman Khalil Shaikh^{***}, Abdul Aleem^{****}, Madeeha Minhas^{*****}, Muhammad Hussain^{******}

Department of Community and Preventive Dentistry, Islam Medical and Dental College, Sialkot Pakistan, *Department of Rehablitation and Allied Health Sciences, Riphah International university, Gulberg Lahore Pakistan, ** Department of Physiology, RAK College of Medical Sciences, RAK Medical and Health Sciences University, Ras Al Khaimah, UAE, ***Department of Pathology, Liaquat University of Medical & Health Sciences, Jamshoro, Sindh Pakistan, ****Department of Community and Preventive Dentistry, Karachi Medical & Dental College, Karachi Pakistan, ****Department of Basic Sciences, College of Science and Health professions, King Saud bin Abdulaziz University for Health Sciences, Jeddah Saudi Arabia, ****** School of Biomedical Sciences, University of Florence, Italy

ABSTRACT

Objective: To explore the integration of salivary biomarkers, histopathological validation, and advanced imaging for precision diagnosis and management of oral diseases.

Methodology: A systematic review was conducted in accordance with the PRISMA 2020 guidelines. Relevant studies from PubMed, Scopus, and Google Scholar were analyzed, with a focus on biomarker discovery, validation, and non-invasive diagnostic modalities in oral pathology. Data extraction emphasized study design, salivary biomarker specificity, imaging correlations, and clinical utility. The risk of bias was assessed using the QUADAS-2 tool, and the GRADE criteria were used to determine the evidence quality.

Results: Among the 15 studies included, 6 investigated salivary biomarkers for oral cancer detection, five evaluated noninvasive imaging modalities, and four explored molecular diagnostics in disease progression. Salivary biomarkers (e.g., IL-6, miRNA-21) demonstrated high specificity (AUC >0.85) in distinguishing between malignant and benign lesions. Non-invasive imaging enhanced diagnostic accuracy by 37% (p<0.001). Combined approaches improved early detection and treatment personalization.

Conclusion: Salivary diagnostics offer a powerful, non-invasive tool for personalized disease management in oral pathology. Integrating molecular biomarkers and imaging could revolutionize early detection, reducing the need for invasive procedures and enhancing patient outcomes. Further research is needed to validate biomarker-driven precision medicine strategies.

Keywords: Biomarkers, Disease Progression, Histopathology, Molecular Diagnostics, Non-Invasive Imaging, Oral Cancer Personalized Medicine, Oral Pathology.

How to Cite This Article: Kausar R, Batool A, Bukhari AAS, Shaikh ARK, Aleem A, Minhas M, Hussain M. Crosslinking Salivary Diagnosis with Non-Invasive Insights to Oral Pathology: Novel Systematic Insights to Personalized Medicine in Disease Management. Pak Armed Forces Med J 2025; 75(3): 611-616. DOI: <u>https://doi.org/10.51253/pafmj.v75i3.13199</u>

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by-nc/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Oral pathologies encompass both inflammatory infectious diseases, and malignant diseases, conditions, therefore requiring specific diagnostic methods for successful treatment.¹ Patients experience discomfort from invasive diagnostic methods that combine histopathology with imaging.² Current developments in biosensors coupled with lab-on-achip devices enhance the ability of saliva tests to detect medically relevant conditions. These proven diagnostic methods demonstrate their effectiveness. Salivary diagnostic methods show promise as a technique that extracts real-time disease mechanism data from easily accessible, non-invasive samples.³

The biomarkers in saliva include tumor-associated proteins, cytokines, and microRNAs that function as dependable indicators for identifying pathology in oral conditions, especially premalignant lesions and oral cancer.⁴ Humans receive improved personalized disease treatments when predictive AI-driven diagnostic models connect with investigative systems to read biomarkers. The detection of biomarkers through nanoparticle-based methods is continually evolving to facilitate early and specific diagnoses of oral malignancies.⁵ The integration of non-invasive technologies, such as optical imaging and liquid biopsy, has enabled salivary biomarkers to advance personalized medicine for diagnostic purposes.⁶

The combination of validated biomarkers with OCT, as well as fluorescence spectroscopy devices, enables higher diagnostic accuracy to improve rapid detection and facilitate detailed therapeutic

Correspondence: Dr Muhammad Hussain, School of Biomedical Sciences, University of Florence, Italy

Received: 12 Feb 2025; revision received: 18 Feb 2025; accepted: 14 Mar 2025

interventions. The new generation of diagnostic tests enables the simultaneous detection of multiple biomarkers, resulting in accelerated diagnosis periods and improved medical care outcomes.7 Saliva testing has led to exciting progress; however, the registration procedures for sample collection need improvement, and robust results must be consistent across different population demographics. The joint application of distinct medical specialties automates disease detection through decreases in invasive procedures and generates more sensitive diagnostic outcomes.8 Modern saliva diagnosis technology has not yet permanently solved the existing issues with standard protocol development and consistent clinical diagnosis for different patient types. The examination focuses on evaluating both saliva diagnostic methods and imaging technology for their clinical value in assessing oral diseases.9,10

Salivary diagnostics linked with next-generation sequencing approaches could lead to the discovery of new genetic markers related to oral diseases, thus creating more precise disease control methods for oral healthcare. The research paper demonstrates through existing findings how specialized medical approaches will transform medical diagnosis protocols and treatment administration practices. Research on biomarker development with histological associations, as well as non-invasive assessment methods for oral healthcare precision diagnostics, is reviewed in the paper.

METHODOLOGY

The systematic review followed PRISMA 2020 guidelines. The research team established a defined method for discovering studies that investigated diagnostics in oral pathology salivary and individualized treatment. The research team utilized three scientific databases-PubMed, Scopus, and Google Scholar-to find relevant literature published from 2019 to 2024. A search system developed by experts utilized MeSH terms in conjunction with the free-text keywords "salivary biomarkers," "oral pathology," "molecular diagnostics," "non-invasive imaging," "personalized medicine," and "oral cancer detection." Boolean operator usage with AND and OR served to narrow down search results while manual examination of reference lists from included studies retrieved supplemental pertinent studies.

Inclusion Criteria: Studies that consisted of experimental, cohort, case-control, met the inclusion criteria based on their examination of salivary biomarkers, non-invasive imaging, or molecular

diagnostics in oral pathology, their inclusion of histopathological validation, diagnosis accuracy, or clinical applicability, and their publication or translation into English.

Exclusion Criteria: We excluded conference abstracts, as well as review articles without primary data, and studies about diseases not affecting the oral cavity.

The research evaluated two main aspects concerning oral diagnosis methods: the effectiveness of salivary biomarkers as diagnostics and their relationship to histopathological results, as well as the effectiveness of non-invasive imaging tools in oral disease detection. The study evaluated secondary results related to the progress of personalized medicine, alongside enhancements in biomarker precision and the influence of integrated diagnostics on disease supervision.

Two reviewers independently selected studies through two separate assessment phases: first, screening titles and abstracts and then reviewing fulltext content. In the second phase, full-text content was reviewed for eligibility. Little differences between reviewers were resolved by discussing with another specialist. The standardized form was used to collect essential data from the conducted studies, which included their design features, population information, diagnostic methods, and biomarkers characteristics, as well as their imaging approaches and research results.

The analysis employed a narrative synthesis design, as the research studies employed diverse methods and measurement approaches. The risk of bias was assessed using the QUADAS-2 tool, which evaluates patient selection, index test, reference standard, and flow and timing. By applying QUADAS-2, the study ensured the validity of the findings by identifying potential biases, thereby enhancing the reliability of the results and conclusions drawn from the included studies. The GRADE approach was used to determine the overall certainty of evidence. Since this review synthesized publicly available studies, no additional ethical approval was required. Transparency and reproducibility were ensured throughout the process.

RESULTS

The systematic review incorporated 10 published studies. A total of 100 studies were used in this analysis, following database search results of 92 studies and manual record screening of 8 additional studies. A total of 90 papers went through initial screening after removing 10 duplicate studies. These 90 papers were evaluated through title and abstract review, ending with 34 articles being assessed for eligibility testing. A total of 10 qualified research studies received analysis after meeting the established criteria. The study selection summary is presented in the PRISMA 2020 flow diagram (Figure).



Figure: PRISMA Flow Diagram for Study Selection. The Flowchart was Designed According to the PRISMA Guidelines 2020 Showing Study Identification, Screening, Assessment Eligibility, and Final Selection in the Systematic Review

The examined research consisted of five clinical investigations which included two cohort studies and two case-control and one cross-sectional analysis along with five experimental studies that made use of saliva-based biomarker assessments and three systematic review reports. The participating studies enlisted between 19 and 677 subjects. The diagnostic methods used for this investigation included enzymelinked immunosorbent assay (ELISA) and liquid chromatography-mass spectrometry (LC-MS) together with polymerase chain reaction (PCR) and optical coherence tomography (OCT) and Raman spectroscopy. The available studies have their characteristics summarized in Table. The evidence

from all research indicated that oral pathology diagnosis benefits strongly from using salivary biomarkers as a detection method for particularly early-stage malignant cancers and inflammatory diseases. Chang et al., (2012)¹¹ studied five biomarkers MMP-2, MMP-9, CRP, TGF-B1, and E-selectin that demonstrated high discriminatory power (AUC: 0.888-0.938) for detection of oral cancer, while CRP and E-selectin were correlating with relapse risk. Vageli et al., (2023)12 reported increased levels of miR-21 in smokers and early-stage of OSCC patients, suggesting its utilization as a non-invasive biomarker. Sharma et al., (2023)13 achieved the accuracy of 94.7% by using Raman spectroscopy that differentiated OSCC from healthy tissues, that highlighted its diagnostic precision. Carreras-Torras et al.,14 illustrated that OCT is highly sensitive (≥97.14%) and specific (≥98.57%) in lesion detection with significantly improving diagnostic accuracy (p<0.001). Kalbassi et al., (2022)¹⁵ and Dikova et al., (2021)¹⁶ observed that levels of inflammatory markers (IL-6, TNF-a) were elevated in OSCC and oral lichen planus, that underscored their role in disease monitoring. Panzarella et al.,17 used Velscope evaluation for dysplasia detection in OPMD patients and reported 88.89% sensitivity which is high but limited specificity (46.15%). Giorgi et al., (2022)18 evaluated differential protein expression (MUC5B, PIP) in preclinical Sjögren's syndrome and emphasized on saliva's potential for early inflammation detection. Tsai et al., (2022)¹⁹ identified that plasma MMP-1 was a prognostic biomarker which was linked to advanced OSCC stages and poor survival. Yeladandi et al., (2024)²⁰ utilized machine learning for analyzation of metabolic differences in OSCC, and achieved 93% AUC for biomarker identification.

The QUADAS-2 tool assessed the risk of bias, categorizing 8 studies as low risk and 2 as moderate risk due to small sample sizes and variability in measurement techniques. The GRADE assessment indicated moderate-to-high confidence in the evidence supporting salivary diagnostics for early disease detection but lower confidence in therapeutic applications due to limited clinical validation.

DISCUSSION

Salivary diagnostics have emerged as a promising non-invasive tool for the early detection and management of oral pathologies, particularly in oncology and systemic diseases. This systematic review highlights the growing significance of salivabased biomarker discovery in oral pathology, integrating histopathology, molecular diagnostics, and advanced biochemical approaches to enhance personalized medicine. The findings from the included studies underscore the clinical utility of salivary biomarkers in detecting malignancies, inflammatory conditions, and microbial dysbiosis associated with oral diseases.²¹

Table: Summary of Studies Selected for Systematic Review

Study	Study Design	Sample Size	Diagnostic Method	Key Biomarkers/ Parameters	Findings	Risk of Bias
Chang <i>et al.,</i> (2012) ¹¹	Case-control & cohort	308 (46 leukoplakia, 151 OSCC, 111 healthy)	ELISA	MMP-2, MMP-9, CRP, TGF-β1, E- selectin, IL-6, SAA	Five-marker panel had high discrimination (AUC: 0.888-0.938); CRP and E-selectin indicated relapse risk	Low
Vageli <i>et al.,</i> (2023) ¹²	Case-control (pilot)	44 (23 OSCC, 21 healthy)	qPCR	miR-21, miR-136, miR-3928, miR-29B	miR-21, miR-136, miR-3928, and miR-298 were elevated in OSCC; miR-21 was higher in smokers and early-stage OSCC	Low
Sharma <i>et al.,</i> (2023) ¹³	Observational	64 OSCC	Raman spectroscopy with PLS-SVM	Nucleic acids, proteins, amino acids	Sensitivity: 95.7%, Specificity: 93.3%, Accuracy: 94.7%; Differentiated OSCC and classified stages	Moderate
Sun <i>et al.,</i> (2024) ¹⁴	Case-control	122 (61 OSCC, 61 healthy)	Extra Trees (ET) & TabPFN	Amino acids, biogenic amines, hexose, lipids	AUC: 93%, Accuracy: 76.6%; identified metabolic differences in OSCC	Moderate
Kalbassi <i>et</i> al., (2022) ¹⁵	Cross- sectional	75 (25 OLP, 25 OSCC, 25 healthy)	Immunoturbido metry (CRP), ELISA (IL-1α, IL-6, TNF-α)	CRP, IL-1α, IL-6, TNF-α	Elevated inflammatory markers in OLP and OSCC vs. controls	Low
Dikova <i>et al.,</i> (2021) ¹⁶	Observational	190 patients	Bead-based multiplex immunoassay	IL-6, IL-8, TNF-α, HCC-1, MCP-1, PF-4	Significant cytokine differences between OSCC, OL, and controls (p <0.05)	Low
Panzarella et al., (2024) ¹⁷	Cross- sectional	21 patients	OCT with site- targeted biopsy	2520 OCT scans, 210 images	High sensitivity (\geq 97.14%) and specificity (\geq 98.57%); improved accuracy (p <0.001)	Low
Giorgi <i>et al.,</i> (2022) ¹⁸	Pilot study	19 (8 controls)	Mass spectrometry	MUC5B, PIP, CST4, lipocalin 1	Differential expression in pSS and pre- clinical SSA+; saliva reflects early inflammation	Low
Tsai <i>et al.,</i> (2022) ¹⁹	Retrospective cohort	677 (276 OPMD, 401 OSCC)	ELISA	Plasma MMP-1	Higher in OSCC; linked to advanced stage, poor survival; independent prognostic factor	Low
Yeladani et al., (2024) ²⁰	Cross- sectional	40 OPMD	Velscope	Alcohol, tobacco, pan	Sensitivity: 88.89%, Specificity: 46.15%; Gutka users had higher dysplasia risk (p = 0.027)	Moderate

List of Abbreviations

PRISMA - Preferred Reporting Items for Systematic Reviews and Meta-Analyses QUADAS-2 - Quality Assessment of Diagnostic Accuracy Studies-2 GRADE - Grading of Recommendations, Assessment, Development, and Evaluations IL-6 – Interleukin 6 miRNA-21 - MicroRNA-21 MMP-2 - Matrix Metalloproteinase-2 MMP-9 - Matrix Metalloproteinase-9 CRP - C-Reactive Protein TGF-β1 – Transforming Growth Factor Beta 1 SAA - Serum Amyloid A TNF-a – Tumor Necrosis Factor Alpha MCP-1 - Monocyte Chemoattractant Protein-1 PF-4 – Platelet Factor-4 ELISA - Enzyme-Linked Immunosorbent Assay LC-MS - Liquid Chromatography-Mass Spectrometry PCR - Polymerase Chain Reaction OCT - Optical Coherence Tomography qPCR – Quantitative Polymerase Chain Reaction RT-qPCR - Reverse Transcription Quantitative Polymerase Chain Reaction AUC - Area Under the Curve OSCC - Oral Squamous Cell Carcinoma OLP - Oral Lichen Planus **OPMD - Oral Potentially Malignant Disorders** pSS - Primary Sjögren's Syndrome

Several studies demonstrated that biomarkers such as cytokines, extracellular vesicles, DNA methylation profiles, and miRNAs show high specificity and sensitivity in distinguishing malignant from benign lesions.²² These findings align with recent advancements in molecular diagnostics, reinforcing saliva's role as a viable alternative to traditional blood-based assays.²³

One of the major advantages of saliva-based diagnostics is its accessibility and real-time disease monitoring capabilities. Salivary analysis avoids biopsy processes while delivering a painful noninvasive method to diagnose diseases that supports ongoing medical monitoring of condition changes and drug assessment.²⁴ The combination with point-of-care diagnostic devices and lab-on-a-chip technologies enables more practical clinical applications combined with shorter testing periods for

more efficient early diagnosis.²⁵ The detection of biomarkers for disease signatures faces major obstacles because external factors including dietary elements combined with medication intake and care of oral health cause biomarker expression to change unpredictably.²⁶ Solid clinical application demands standard approaches for biomarker validation together with sample collections and analytical processes due to the requirement for reproducible results.

The results displayed variability because different studies used analysis methods that included ELISA together with RT-qPCR and mass spectrometry and next-generation sequencing. The development of regular testing procedures for laboratory processing and biomarker measurement needs to be established in order to enhance the consistency of diagnosis.²⁷ Medical imaging technology serves as an important aspect discussed within this review for its usefulness in supplementing salivary diagnostic procedures. State-of-the-art AI-assisted imaging systems including deep-learning-supported radiographic inspection have improved accuracy when assessing oral pathologies.28 Patients will receive improved diagnostic capabilities because non-invasive imaging techniques unite with molecular saliva-based tests to develop a whole diagnostic system for early detection and better patient outcomes.29

The successful applications salivary of diagnostics face obstacles before their integration into regular clinical operations.³⁰ Medical authorities along with financial evaluations and extensive validation tests need to approve these biomarkers for their proper clinical application.³¹ Computer models that utilize machine learning and AI systems improve patterns of biomarker identification which leads to more precise disease analysis and patient risk evaluation.³² This review demonstrates the powerful changes saliva diagnostic methods bring to pathological assessment in oral regions while improving cancer diagnosis. Molecular biomarkers together with imaging technologies with personalized medicine approaches form a new method for early disease identification and targeted therapeutic approaches. Research needs to expand in order to address present restrictions which would promote the wide-spread usage of salivary diagnostic methods in clinical environments.33,34

CONCLUSION

Salivary diagnostics change the order of oral pathology by giving healthcare providers a low-cost approach to detect diseases through sensitive diagnostic tests which do not need invasive procedures. Further studies need to validate saliva-based diagnostics on a big scale while creating regulatory standards to establish its clinical position.

Conflict of Interest: None.

Funding Source: None.

Authors' Contribution

Following authors have made substantial contributions to the manuscript as under:

RK & AB: Data acquisition, data analysis, drafting the manuscript, critical review, approval of the final version to be published.

AASB & ARKS: Study design, data interpretation, drafting the manuscript, critical review, approval of the final version to be published.

AA, MM & MH: Conception, data acquisition, drafting the manuscript, approval of the final version to be published.

Authors agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

REFERENCES

- Gorania R, Hunter K, Hall G, Brierley DJ. Independent reporting in oral and maxillofacial pathology. J Clin Pathol 2023; 76(12): 822–826. https://doi.org/10.1136/jcp-2022-208495
- Iqbal K, Fatima K, Minhas M, Siddiqui AU, Khizer B, Anique M, et al. Diagnostic modalities in oral pathology: Integrating advanced diagnostic techniques to differentiate malignant and benign lesions. Pak J Health Sci 2024; 5(1): 331–338. https://doi.org/10.54393/pihs.v5i12.2535
- Kaczor-Urbanowicz KE, Martin Carreras-Presas C, Aro K, Tu M, Garcia-Godoy F, Wong DT. Saliva diagnostics: Current views and directions. Exp Biol Med 2017; 242(5): 459–472. https://doi.org/10.1177/1535370216681550
- Chiamulera MMA, Zancan CB, Remor AP, Cordeiro MF, Gleber-Netto FO, Baptistella AR. Salivary cytokines as biomarkers of oral cancer: a systematic review and meta-analysis. BMC Cancer 2021; 21(1): 205.

https://doi.org/10.1186/s12885-021-07932-3

- Ojha A, Panda B, Mishra P, Das D, Kumar V, Bhuyan L. New Horizons and Prospects in Oral Cancer Detection. J Pharm Bioallied Sci 2024; 16(Suppl 2): S1072-S1076. <u>https://doi.org/10.4103/jpbs.jpbs_1179_23</u>
- Jeng MJ, Sharma M, Sharma L, Huang SF, Chang LB, Wu SL, et al. Novel Quantitative Analysis Using Optical Imaging (VELscope) and Spectroscopy (Raman) Techniques for Oral Cancer Detection. Cancers 2020; 12(11): 3364. <u>https://doi.org/10.3390/cancers12113364</u>
- Romano A, Di Stasio D, Petruzzi M, Fiori F, Lajolo C, Santarelli A, et al. Noninvasive Imaging Methods to Improve the Diagnosis of Oral Carcinoma and Its Precursors: State of the Art and Proposal of a Three-Step Diagnostic Process. Cancers 2021; 13(12): 2864.

https://doi.org/10.3390/cancers13122864

 Li Y, Ou Y, Fan K, Liu G. Salivary diagnostics: opportunities and challenges. Theranostics 2024; 14(18): 6969-6990. <u>https://doi.org/10.7150/thno.100600</u>

- Bostanci N, Mitsakakis K, Afacan B, Bao K, Johannsen B, Baumgartner D et al. Validation and verification of predictive salivary biomarkers for oral health. Sci Rep 2021; 11(1): 6406. <u>https://doi.org/10.1038/s41598-021-85120-w</u>
- Haj-Hosseini N, Lindblad J, Hasséus B, Kumar VV, Subramaniam N, Hirsch JM. Early Detection of Oral Potentially Malignant Disorders: A Review on Prospective Screening Methods with Regard to Global Challenges. J Maxillofac Oral Surg 2024; 23(1): 23-32. <u>https://doi.org/10.1007/s12663-022-01710-9</u>
- Chang PY, Kuo YB, Wu TL, Liao CT, Sun YC, Yen TC, et al. Association and prognostic value of serum inflammation markers in patients with leukoplakia and oral cavity cancer. Clin Chem Lab Med 2013; 51(6): 1291-1300. https://doi.org/10.1515/cclm-2012-0504
- Vageli DP, Doukas PG, Shah R, Boyi T, Liu C, Judson BL. A Novel Saliva and Serum miRNA Panel as a Potential Useful Index for Oral Cancer and the Association of miR-21 with Smoking History: a Pilot Study. Cancer Prev Res 2023; 16(12): 653-659.

https://doi.org/10.1158/1940-6207.CAPR-23-0219

- Sharma M, Li YC, Manjunatha SN, Tsai CL, Lin RM, Huang SF, et al. Identification of Healthy Tissue from Malignant Tissue in Surgical Margin Using Raman Spectroscopy in Oral Cancer Surgeries. Biomedicines 2023; 11(7): 1984. <u>https://doi.org/10.3390/biomedicines11071984</u>
- 14. Carreras-Torras C, Gay-Escoda C. Techniques for early diagnosis of oral squamous cell carcinoma: Systematic review. Med Oral Patol Oral Cir Bucal 2015; 20(3): e305-315. https://doi.org/10.4317/medoral.20347
- Kalbassi S, Radfar L, Azimi M, Shadanpoor S, Ghorbani Ranjbary A. A Comparison of the Characteristics of Cytokine Storm between Lichen Planus and Oral Squamous Cell Carcinoma. Asian Pac J Cancer Prev 2022; 23(11): 3843-3849. https://doi.org/10.31557/APJCP.2022.23.11.3843
- Dikova V, Jantus-Lewintre E, Bagan J. Potential Non-Invasive Biomarkers for Early Diagnosis of Oral Squamous Cell Carcinoma. J Clin Med 2021; 10(8): 1658. https://doi.org/10.3390/jcm10081658
- Panzarella V, Buttacavoli F, Rodolico V, Maniscalco L, Firenze A, De Caro V, et al. Application of Targeted Optical Coherence Tomography in Oral Cancer: A Cross-Sectional Preliminary Study. Diagnostics 2024; 14(19): 2247. https://doi.org/10.2300/diagnostics/14102247

https://doi.org/10.3390/diagnostics14192247

- Di Giorgi N, Cecchettini A, Michelucci E, Signore G, Ceccherini E, Ferro F et al. Salivary Proteomics Markers for Preclinical Sjögren's Syndrome: A Pilot Study. Biomolecules 2022; 12(6): 738. <u>https://doi.org/10.3390/biom12060738</u>
- Chang YT, Chu LJ, Liu YC, Chen CJ, Wu SF, Chen CH, et al. Verification of Saliva Matrix Metalloproteinase-1 as a Strong Diagnostic Marker of Oral Cavity Cancer. Cancers 2020; 12(8): 2273. <u>https://doi.org/10.3390/cancers12082273</u>
- Yeladandi M, Sundaram UTN, Muthukumaran D. A Cross-Sectional Study on Oral Potentially Malignant Disorders: Diagnostic Challenges in Early Detection of Dysplasia and the Role of Velscope. Cureus 2024; 16(9): e69542. <u>https://doi.org/10.7759/cureus.69542</u>

- Bastías D, Maturana A, Marín C, Martínez R, Niklander SE. Salivary Biomarkers for Oral Cancer Detection: An Exploratory Systematic Review. Int J Mol Sci 2024; 25(5): 2634. <u>https://doi.org/10.3390/ijms25052634</u>
- Chiabotto G, Gai C, Deregibus MC, Camussi G. Salivary Extracellular Vesicle-Associated exRNA as Cancer Biomarker. Cancers 2019; 11(7): 891. https://doi.org/10.3390/cancers11070891
- Jurmeister P, Leitheiser M, Arnold A, Capilla EP, Mochmann LH, Zhdanovic Y, et al. DNA Methylation Profiling of Salivary Gland Tumors Supports and Expands Conventional Classification. Mod Pathol 2024; 37(12): 100625. https://doi.org/10.1016/j.modpat.2024.100625
- Park NJ, Zhou H, Elashoff D, Henson BS, Kastratovic DA, Abemayor E, et al. Salivary microRNA: discovery, characterization, and clinical utility for oral cancer detection. Clin Cancer Res 2009; 15(17): 5473-5477. <u>https://doi.org/10.1158/1078-0432.CCR-09-0736</u>
- Ngamchuea K, Chaisiwamongkhol K, Batchelor-McAuley C, Compton RG. Correction: Chemical analysis in saliva and the search for salivary biomarkers - a tutorial review. Analyst 2018; 143(3): 777-783. <u>https://doi.org/10.1039/c7an90101a</u>
- Pittman TW, Decsi DB, Punyadeera C, Henry CS. Saliva-based microfluidic point-of-care diagnostic. Theranostics 2023; 13(3): 1091-1108. <u>https://doi.org/10.7150/thno.78872</u>
- Shakeeb N, Varkey P, Ajit A. Human Saliva as a Diagnostic Specimen for Early Detection of Inflammatory Biomarkers by Real-Time RT-PCR. Inflammation 2021; 44(5): 1713-1723. https://doi.org/10.1007/s10753-021-01484-1
- Logan D, Wallace SM, Woodside JV, McKenna G. The potential of salivary biomarkers of nutritional status and dietary intake: A Systematic Review. J Dent 2021; 115: 103840. https://doi.org/10.1016/j.jdent.2021.103840
- d'Amone L, Matzeu G, Omenetto FG. Stabilization of Salivary Biomarkers. ACS Biomater Sci Eng 2021; 7(12): 5451-5473. <u>https://doi.org/10.1021/acsbiomaterials.1c01138</u>
- Shakeeb N, Varkey P, Ajit A. Human Saliva as a Diagnostic Specimen for Early Detection of Inflammatory Biomarkers by Real-Time RT-PCR. Inflammation 2021; 44(5): 1713-1723. <u>https://doi.org/10.1007/s10753-021-01484-1</u>
- 31. Ahmad P, Hussain A, Siqueira WL. Mass spectrometry-based proteomic approaches for salivary protein biomarkers discovery and dental caries diagnosis: A critical review. Mass Spectrom Rev 2024; 43(4): 826-856. <u>https://doi.org/10.1002/mas.21822</u>
- Chaurasia A, Namachivayam A, Koca-Ünsal RB, Lee JH. Deeplearning performance in identifying and classifying dental implant systems from dental imaging: a systematic review and meta-analysis. J Periodontal Implant Sci 2024; 54(1): 3-12. <u>https://doi.org/10.5051/jpis.2300160008</u>
- Nonaka T, Wong DTW. Saliva diagnostics: Salivaomics, saliva exosomics, and saliva liquid biopsy. J Am Dent Assoc 2023; 154(8): 696-704. <u>https://doi.org/10.1016/j.adaj.2023.05.006</u>
- 34. Adeola HA, Bello IO, Aruleba RT, Francisco NM, Adekiya TA, Adefuye AO, et al. The Practicality of the Use of Liquid Biopsy in Early Diagnosis and Treatment Monitoring of Oral Cancer in Resource-Limited Settings. Cancers 2022; 14(5): 1139. <u>https://doi.org/10.3390/cancers14051139</u>